

# Jamming signatures observed in peptide translocation

Jason T. Deckman and Mahua Roy  
*Department of Chemistry, University of California, Irvine*

Ioan Andricioaei\*  
*Department of Chemistry, University of California, Irvine, CA 92697*  
\*Email: andricio@uci.edu, Tel.: 949-824-3569.  
(Dated: November 8, 2013)

Upon peptide passage through membrane pores, peptides undergo conformational transitions and sample intermediates that block the transmembrane current [1–8] that would otherwise flow in an open pore under a potential drop. These intermediate states can be considered jammed states. On the other hand, the phenomenology of jamming has been widely studied, separately in the context of dynamical arrest of macroscopic granular matter and in that of macroscopic glasses [9–14]. Jamming can be identified by signature characteristics in the force distributions,  $p(f)$  - the force profile of a jammed system will exhibit a peak in  $p(f)$ , that deviates from the exponential decay in the forces of an unjammed state [15–20]. In previous work, we have applied such jamming analyses to study protein folding in aqueous solution [21]. Herein we investigate the force distributions associated with an unstructured peptide, CAMA [22, 23] both in free state and within the constriction of a pore lumen.

Translocation of an unstructured peptide, **Cecropin A - MAgainin(CAMA)** with a sequence of 20 amino acids ( $KWKLFFKKIGIGKFLQSAKKF - NH_2$ ), through biological nano-pore  $\alpha$ -HL [24], was studied using single molecule electrophysiology, inserted from the trans-end [25, 26]. Molecular dynamics simulations of the above process at complete atomistic resolution [27–32], was successful in capturing the sub-states, which induce partial or complete blockage of the pore, reflected as spikes in the current blockage signature. Three states of CAMA are chosen. First is the free peptide, which assumes a heterogeneous kinked shape due the presence of two alternate glycines(G) at positions 9 and 11 separated by one Isoleucine (I) at position 10, most reminiscent of an un-jammed peptide. Second, in which the peptide is unfolded within the pore constriction and translocated through the beta-barrel of  $\alpha$ -HL in a linear fashion (less jammed), and lastly where the CAMA traverses the pore in a wound, condensed state, a configuration most akin to a jammed peptide. Representative snapshots of the conformational ensemble assumed at each state are depicted with their respective non-bonded force profiles in Figure 1. All three states exhibit jamming signatures, as marked by the pronounced peak above the mean force, though this peak is shifted more to the right the more unjammed the peptide, *e.g.* the free CAMA, which is shifted farthest. Thus we may surmise that the position of the peaks depends upon the relative degree of jamming between the configurations. Note that the simulation for the free CAMA was carried out at room temperature where jamming is shown, from our previous study, to be extant in peptides. A simulation at a much higher temperature will most likely wash out the jamming traits in the force profile, which can be investigated in a future study.

Translocation trajectories of the peptide traversing the pore in condensed and linear state as well as the dynamics of the free peptide were generated using the CHARMM-27 force field in ACE implicit solvent model (continuum with dielectric constant of 80) using Langevin dynamics (friction coefficient  $10 \text{ ps}^{-1}$ ) at 300K. Translocation is effected by using an accelerated electric field applied to the peptide only, calculated from the applied trans-membrane voltage along the z-axis, given by,  $V = -EL_z$ . The non-bonded or van der Waals interactions were computed using switching function with a cutoff of  $12\text{\AA}$ . The SHAKE algorithm was used to constrain the hydrogen bonds permitting a time step of 2 fs. Unfolding of the peptides within the pore constriction was effected by adding a half quadratic perturbation term to the Hamiltonian, dependent on both time and reaction coordinate, forcing the peptide to move away from its initial configuration using a half harmonic force constant through the use of the biased MD method [33, 34] in CHARMM simulation package.

- 
- [1] Kevin J. Freedman, S. Raza Haq, Joshua B. Edel, Per Jemth, and Min Jun Kim. Single molecule unfolding and stretching of protein domains inside a solid-state nanopore by electric field. *Sci. Rep.*, 3, 2013.
  - [2] Cees Dekker. Solid-state nanopores. *Nat Nano*, 2:209–215, 2013.
  - [3] Linda Payet, Marlene Martinho, Manuela Pastoriza-Gallego, Jean-Michel Betton, Loc Auvray, Juan Pelta, and Jrme Math. Thermal unfolding of proteins probed at the single molecule level using nanopores. *Analytical Chemistry*, 84(9):4071–4076,

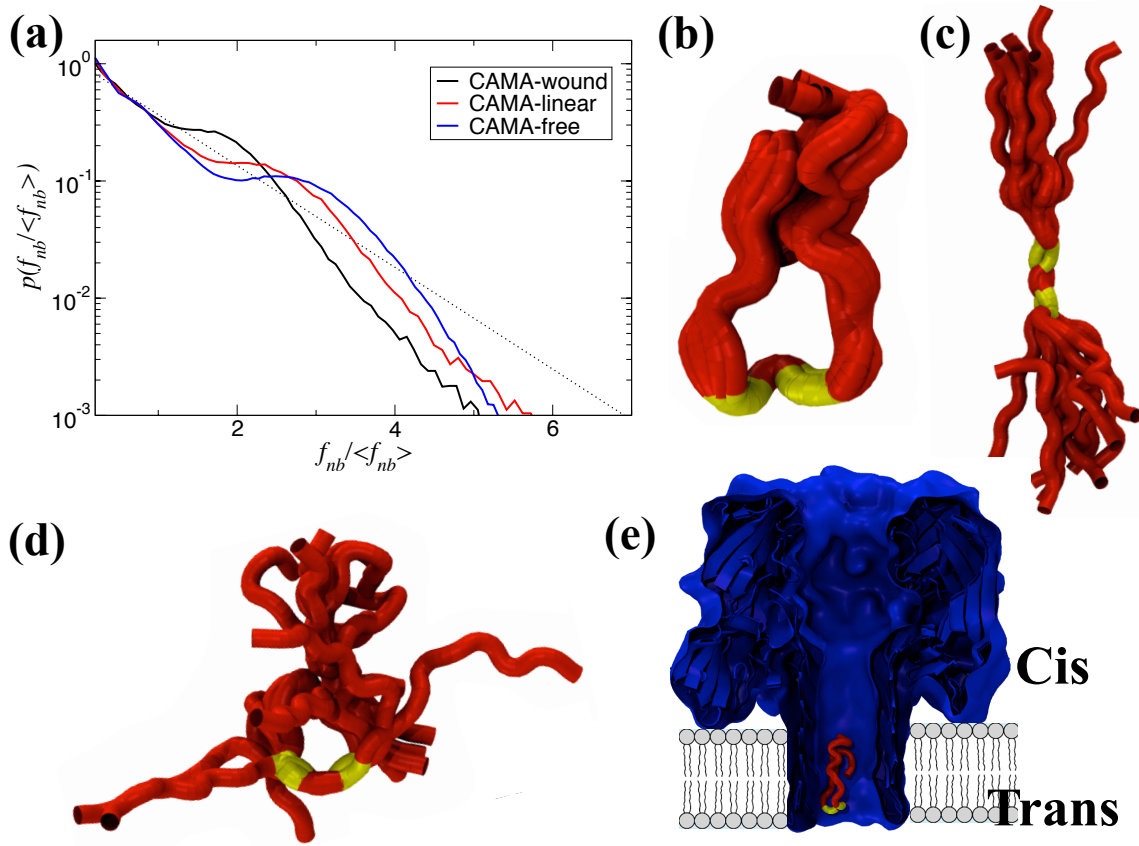


FIG. 1: (a) The non-bonded, normalized force distributions for three different states of CAMA, ranging from jammed (wound) to marginally jammed to relatively un-jammed (free). All three exhibit jamming signatures, namely a pronounced peak about the exponential line (dashed), though this is shifted to higher values the more apparently un-jammed the peptide. Additionally, the tails of those that are less jammed are closer to the exponential line. Conformational ensembles of CAMA P6 in (b) wound state (c) Linear and (d) free state. (e) Translocation of peptide CAMA - P6 through lumen of  $\alpha$ -hemolysin

- 2012.
- [4] L. Movileanu, S. Howorka, O. Braha, and H. Bayley. Detecting protein analytes that modulate transmembrane movement of a polymer chain within a single protein pore. *Nature biotechnology*, 18(10):1091–1095, October 2000.
  - [5] G. Oukhaled, J. Mathé, A.-L. Biance, L. Bacri, J.-M. Betton, D. Lairez, J. Pelta, and L. Auvray. Unfolding of proteins and long transient conformations detected by single nanopore recording. *Phys. Rev. Lett.*, 98:158101, Apr 2007.
  - [6] Manuela Pastoriza-Gallego, Leila Rabah, Gabriel Gibrat, Bndicte Thiebot, Franoise Gisou van der Goot, Loc Auvray, Jean-Michel Betton, and Juan Pelta. Dynamics of unfolded protein transport through an aerolysin pore. *Journal of the American Chemical Society*, 133(9):2923–2931, 2011.
  - [7] Radu Stefureac, Landon Waldner, Peter Howard, and JeremyS. Lee. Nanopore analysis of a small 86-residue protein. *Small*, 4(1):59–63, 2008.
  - [8] Mohammad Mohammad and Liviu Movileanu. Excursion of a single polypeptide into a protein pore: simple physics, but complicated biology. *European Biophysics Journal*, 37:913–925, 2008. 10.1007/s00249-008-0309-9.
  - [9] C. Heussinger and J.-L. Barrat. Jamming transition as probed by quasistatic shear flow. *Phys. Rev. Lett.*, 102:218303, 2009.
  - [10] D. Head. Critical scaling and aging in cooling systems near the jamming transition. *Phys. Rev. Lett.*, 102:138001, 2009.
  - [11] P. Olsson and S. Teitel. Critical scaling of shear viscosity at the jamming transition. *Phys. Rev. Lett.*, 99:178001, 2007.
  - [12] D. Vagberg, D. Valdez-Balderas, M.A. Moore, P. Olsson, and S. Teitel. Finite-size scaling at the jamming transition: Corrections to scaling and the correlation-length critical exponent. *Physical Review E*, 83:030303(R), 2011.
  - [13] L.E. Silbert. Jamming of frictional spheres and random loose packing. *Soft Matter*, 6:2918–2924, 2010.
  - [14] J.H. Snoeijer, T.J.H. Vlugt, W.G. Ellenbroek, M. van Hecke, and J.M.J. van Leeuwen. Ensemble theory for force networks in hyperstatic granular matter. *Phys. Rev. E*, 70:061306, Dec 2004.
  - [15] J.H. Snoeijer, T.J.H. Vlugt, M. van Hecke, and W. van Saarloos. Force network ensemble: A new approach to static granular matter. *Phys. Rev. Lett.*, 92:054302, 2004.
  - [16] B.P. Tighe, J.E.S. Socolar, D.G. Schaeffer, W.G. Mitchener, and M.L. Huber. Force distributions in a triangular lattice of

- rigid bars. *Phys. Rev. E*, 72:031306, 2005.
- [17] B. P. Tighe, J. H. Snoeijer, T. J. H. Vlugt, and M. van Hecke. The force network ensemble for granular packings. *Soft Matter*, 6:2908–2917, 2010.
  - [18] J.A. Dijksman, G.H. Wortel, L.T.H. van Dellen, O. Dauchot, and M. van Hecke. Jamming, yielding, and rheology of weakly vibrated granular media. *Phys. Rev. Lett.*, 107:108303, Sep 2011.
  - [19] P. M. Reis, R. A. Ingale, and M. D. Shattuck. Caging dynamics in a granular fluid. *Phys. Rev. Lett.*, 98:188301, 2007.
  - [20] D. Bi, J. Zhang, B. Chakraborty, and R. P. Behringer. Jamming by shear. *Nature*, 480(7377):355–358, 2011.
  - [21] Prasanth P Jose and Ioan Andricioaei. Similarities between protein folding and granular jamming. *Nat Commun.*, 3:1161–6170, 2012.
  - [22] Alina Asandei, Aurelia Apetrei, Yoonkyung Park, Kyung-Soo Hahm, and Tudor Luchian. Investigation of single-molecule kinetics mediated by weak hydrogen bonds within a biological nanopore. *Langmuir*, 27(1):19–24, 2011.
  - [23] Loredana Mereuta, Irina Schiopu, Alina Asandei, Yoonkyung Park, Kyung-Soo Hahm, and Tudor Luchian. Protein nanopore-based, single-molecule exploration of copper binding to an antimicrobial-derived, histidine-containing chimera peptide. *Langmuir*, 28(49):17079–17091, 2012.
  - [24] Langzhou Song, Michael R. Hobaugh, Christopher Shustak, Stephen Cheley, Hagan Bayley, and J. Eric Gouaux. Structure of staphylococcal  $\alpha$ -hemolysin, a heptameric transmembrane pore. *Science*, 274(5294):1859–1865, 1996.
  - [25] Oleg V. Krasilnikov, Petr G. Merzlyak, Liliya N. Yuldasheva, Cludio G. Rodrigues, Sucharit Bhakdi, and Angela Valeva. Electrophysiological evidence for heptameric stoichiometry of ion channels formed by staphylococcus aureus alpha-toxin in planar lipid bilayers. *Molecular Microbiology*, 37(6):1372–1378, 2000.
  - [26] L. Mereuta, M. Roy, A. Asandei, J. K. Lee, Y.Park, I. Andricioaei, and T. Luchian. Slowing down single-molecule trafficking through a protein nano-pore reveals intermediates for peptide translocation. *Nature Scientific Reports [under Review]*, 2013.
  - [27] Aleksij Aksimentiev and Klaus Schulten. Imaging  $\alpha$ -hemolysin with molecular dynamics: Ionic conductance, osmotic permeability, and the electrostatic potential map. *Biophysical Journal*, 88(6):3745 – 3761, 2005.
  - [28] Aleksei Aksimentiev. Deciphering ionic current signatures of dna transport through a nanopore. *Nanoscale*, 2:468–483, 2010.
  - [29] Pu Tian and Ioan Andricioaei. Repetitive pulling catalyzes co-translocational unfolding of barnase during import through a mitochondrial pore. *Journal of Molecular Biology*, 350(5):1017 – 1034, 2005.
  - [30] Tom Z Butler, Jens H Gundlach, and Mark Troll. Ionic current blockades from DNA and RNA molecules in the alpha-hemolysin nanopore. *Biophysical journal*, 93(9):3229–3240, November 2007.
  - [31] Yacov Kantor and Mehran Kardar. Anomalous dynamics of forced translocation. *Phys. Rev. E*, 69:021806, Feb 2004.
  - [32] Tom Z. Butler, Jens H. Gundlach, and Mark A. Troll. Determination of rna orientation during translocation through a biological nanopore. *Biophysical Journal*, 90(1):190 – 199, 2006.
  - [33] Emanuele Paci and Martin Karplus. Forced unfolding of fibronectin type 3 modules: an analysis by biased molecular dynamics simulations. *Journal of Molecular Biology*, 288(3):441 – 459, 1999.
  - [34] Pu Tian and Harris D. Bernstein. Molecular basis for the structural stability of an enclosed  $\beta$ -barrel loop. *Journal of Molecular Biology*, 402(2):475 – 489, 2010.